

Risk factors and physiopathology of candidiasis

Jean-Marcel Senet

Laboratoire d'Immunologie, Faculté de Pharmacie, Angers, France

- Summarv Recent epidemiological surveys have demonstrated an important increase in nosocomial infections among which Candida sp. plays an increasingly prominent role. Candida is now involved in about 10% of all septicemia and leads to a very high mortality rate in immunodepressed patients. Clinical studies show that any modification of the host immune status can facilitate the proliferation of endogenous Candida which, according to the importance of the immune deficiency, can provoke diseases ranging from benign localized mucocutaneous candidosis to sometimes lethal systemic invasions. The pathogenic behavior of *Candida* cells is mainly due to a very high phenotypic biodiversity. Following even very slight environmental modifications, it may change its behavior through the appearance of new or amplified properties such as tube formation, adherence, protease secretion, etc. Together with the impairment of host defenses, these new invasive properties lead to the so-called opportunistic pathogenicity of *Candida* cells. From a host point of view, after the integrity of surface teguments, the mucosal protection is ensured by the Th1 "cellular" immune response which, through pro-inflammatory cytokine production, boosts the efficacy of the phagocytes (Polymorphonuclear cells and macrophages). Neutrophils are of particular importance as deep seated Candida proliferation is mostly associated with neutropenia. Whatever the pathogenic process, it is mostly due to modifications provoked by increasing medical awareness which makes patients more susceptible to illness. A better knowledge of the precise mechanisms involved and twould lead to improved strategies for prevention.
- Key words Candida, Candidiasis, Pathophysiology, Opportunistic disease, Epidemiology, Immune deficiency

Factores de riesgo y fisiopatología de la candidiasis

Resumen Estudios epidemiológicos recientes han demostrado un importante aumento de las infecciones nosocomiales, entre las que Candida juega un papel cada vez más relevante. Candida se encuentra implicada en el 10% de las septicemias y provoca una mortalidad elevada en pacientes inmunosuprimidos. Los estudios clínicos muestran que cualquier modificación del estado inmunológico del huésped puede facilitar la proliferación endógena de Candida que, dependiendo del alcance de la inmunodeficiencia, puede provocar enfermedades que van desde candidiasis mucocutáneas localizadas benignas hasta invasiones sistémicas, en ocasiones letales. El comportamiento patógeno de Candida se debe principalmente a una biodiversidad fenotípica elevada. Incluso modificaciones ambientales mínimas pueden cambiar su comportamiento mediante la aparición de propiedades nuevas y amplificadas, como la formación de tubos germinales, la adhesión, la secreción de proteasa, etc. Estas nuevas propiedades invasivas unidas a la alteración de las defensas del huésped, provocan una patogenicidad oportunista de Candida. Desde el punto de vista del huésped, junto con la integridad de las barreras superficiales, la protección de las mucosas está asegurada por la respuesta inmune celular de tipo Th1 que, a través de la producción de citocinas pro-inflamatorias, mejora la eficacia de los fagocitos (células polimorfonucleares y macrófagos). Los neutrófilos son particularmente importantes ya que la proliferación de Candida en los tejidos está principalmente asociada con la neutropenia. Cualquiera que sea el proceso patogénico, éste se debe fundamentalmente a las modificaciones asociadas con los avances médicos que hace a los pacientes más susceptibles. Un mejor conocimiento de los mecanismos concretos de patogenicidad implicados debería orientarnos a mejorar las estrategias de prevención.

Palabras clave

candida, Candidiasis, Patofisiología, Infección oportunista, Epidemiología, Inmunodeficiencia

Multidisciplinary approaches to the physio-pathology of candidiasis has produced some new insights.

Three levels of investigation may now be usefully considered :

(i) Epidemiological surveys: this aspect of the relationship between candidal and human populations gives the real importance of the disease in terms of not only quantity but also in terms of evolutivity. Increasingly accurate epidemiological surveys have produced numerous results leading to a better understanding of the multifactorial aspect of candidiasis and to an improved evaluation of prognosis.

(ii) Clinical aspects of candidiasis: increase in medical awareness has led to the identification of new diseases and nosocomial infections among which candidiasis is prominent. Defense impairments provoked by sophisticated therapies and many other iatrogenic comportments are unfortunately taken advantage of by various opportunistic diseases.

(iii) Cellular and molecular biology: recent developments in these fields provide considerable insight into these phenomena. The genetic and phenotypic variability of the *Candida* cell is better understood and the comprehension of the many mechanisms, cells and molecules involved in the immunological discrimination processes of the human host increases day by day.

The physiopathology of candidiasis requires that all these findings be considered within a dynamic context, whatever the epidemiological, clinical, cellular or molecular levels encompassed.

EPIDEMIOLOGICAL FACTORS

From January 1980 through April 1990, 27,200 fungal isolates causing nosocomial infections were reported from 180 hospitals participating in the US National Nosocomial Infections Surveillance (NNIS) system; *Candida* species accounted for 19,621 (72.1%) of these isolates; the rate of candidal bloodstream infection has increased by as much as 487% throughout this decade [1,2]. Other epidemiological surveys note an 11-fold increase during the same period [3]. The incidence of deep-seated mycoses in the northern hemisphere is estimated to be 600 mycosis situations per million population per year [4]. *Candida* sp. jumped from the 8th to the 4th place in septicemia during the period 1984-88 and corresponds to more than 10% of nosocomial septicemia [5].

Thirty-seven reports, published between 1952 and 1992 describing 1,591 cases of systemic *Candida* infections among patients with cancer, showed that species other than *Candida albicans* accounted for 46% of all systemic infections; in particular, *Candida tropicalis* accounted for 25%, *Candida glabrata* for 8%, *Candida parapsilosis* for 7% and *Candida krusei* for 4% [6]. Emerging pathogens such as *Malassezia furfur; Trichosporon beigelii, Rhodotorula* species, *Hansenula anomala, Candida lusitaniae*, were also observed. Organisms once considered environmental contaminants or only industrially important, such as *Candida utilis* and *Candida lipolytica*, have now been implicated as agents of fungemia, onychomycosis, and systemic diseases.

The noticeable increase in *Candida* infection in immuno-compromised patients occurs with a very high mortality rate which sometime reaches 50% [7-9].

According to their variable susceptibility to antifungal agents [10,11], the time needed for an accurate identification, and the life-threatening infections provoked, these new pathogens, which are mainly of iatrogenic origin, increase the burden of clinical mycology

[12,13].

Another important point concerns the origin of the pathogenic strains. For a long time the endogenous commensal situation of the fungus was considered to be the essential source of contamination. Development of accurate DNA typing methods shows that nosocomial yeast infections can sometimes behave like minor epidemics resulting from the selection of more virulent strains [14-17].

FROM A CLINICAL POINT OF VIEW

Candida albicans may be considered as a commensal of the normal flora of the digestive tract. Pathogenicity results essentially from modifications of the host defense mechanisms which secondarily initiate transformations in the fungal behaviour.

The pathogenic behaviour of *Candida* may appear following even a slight modification of the host. Mucocutaneous candidiasis has been observed in people with physiological cellular immune deficiencies. Thrush in the newborn or the elderly may be related to an inefficiency of the thymus. Candida vulvovaginitis associated with pregnancy or use of contraceptives may be linked to the role of progesterone on T-cells and on PMN anti-candidal activity [18-19]. C. albicans possesses an oestrogen-binding protein that links to oestrogens with high affinity and specificity [20]. Moreover, human chorionic gonadotropin (hCG), human luteinizing hormone (hLH), and other CG-like proteins induce the transition of *C. albicans* from the blastospore to the hyphal form [21]. Moreover, clinical information dating back many years suggests that Th1 responses are weakened systematically during pregnancy.

Stress is an often forgotten cause of temporary immunodeficiency. Neuroendocrine regulation and chronobiological effects may notably modulate the immune system and provide the opportunity of fungal proliferation [22].

As Wilson (1962) put it:"*C. albicans* is a better clinician and can discover abnormalities in persons much earlier in the course of the development of such abnormalities than we can with our chemical tests".

In addition to physiological modifications, there is a long list of diseases that may facilitate the development of opportunistic pathogens. Primary or secondary deficits affecting myeloid or lymphoid lineages show the fundamental role of these cells in the control of self-discrimination and homeostasis. Neutropenia and its duration is obviously one of the main causes of systemic *Candida*proliferation [23-25], since mucocutaneous candidiasis is directly related to T-cell deficiencies. Diabetes and other endocrinopathies are also sources of mucocutaneous candidiasis [26,27].

The considerable increase of *Candida* infections over the last decade, is certainly linked to the development of new drugs and techniques which allow the physician to penetrate deeper into the tissular, cellular and molecular intimacy of patients thus opening breaches for *Candida* to invade. These so-called iatrogenic factors involve new chemical and physical therapeutic techniques such as:

- Antibiotherapy, particularly poly-antibiotherapy causes modifications of the mucosal flora leading to proliferation of *Candida* cells [28-31].

- Corticotherapy, directly [32] or through modification of the cytokine network [33] may affect polymorphonulear (PMN) [34], macrophage [35] and T cell activity leading to an impairment of their antifungal activity

[36,37].

- Chemotherapy leading to depletion of leucocytes provides a breakthrough which facilitates fungal infections [38,39]. Moreover chemotherapy may alter PMN function [40] and ulcerate digestive mucosa contributing to proliferation of *Candida* cells [41,42]; these phenomena may be potentiated by anti-acid treatments [43].

- Surgery, principally of the gastro intestinal tract, associated with chemical, physical and psychological stresses favors fungal development [44].

- Catheterism may provoke an injury of the teguments, and also provide a substrate which, once within the blood, is quickly covered by fibronectin, fibrinogen, platelets and other plasma components; it constitutes a support for settlement and proliferation of microorganisms which are, at this level, more resistant to antibiotics and to antifungal agents [45-48].

- Transplantation by itself integrates all iatrogenic factors [49,50].

Most of these iatrogenic factors are related to an impairment of the surface integrity and immune defenses. Increased medical awareness may explain the expanding rate of these diseases as well as the transformation of the spectrum of nococomial pathogens.

CELLULAR AND MOLECULAR ASPECTS

As a diploid eukaryotic cell Candida obviously possesses enough genomic resources to cope with many kinds of environment. From a very simple culture medium providing basic carbon and nitrogen sources at room temperature through to the complexity of the human body, Candida is capable of growing, mutiplying and colonizing. Devoid of sexual reproduction, C. albicans can face the numerous environmental selection factors by important phenotype and karyotype variations, the latest being mainly translocation, chromosome fragmentation and aneuploidy. Karyotype analysis has gained in interest since the methodologies now used offer greater discrimination. However, until recently, their main use has been epidemiologic since it is difficult to establish obvious links between karyotypes and phenotypes. Gene cloning, gene disruption, gene mapping and other molecular genetic techniques have yielded considerable information in the past few years and should certainly prove helpful for the comprehension of the molecular mechanisms involved in candidal variability and virulence.

At the cellular and molecular level, the "commensal" *Candida*, adopting pathogenic behaviour has to switch from a quiet way of life to a more complicated one to overcome the numerous barriers naturally developed by the host. It has to go through many steps to colonize and to proliferate through what is a usually well protected citadel. Decrease in host defenses is not sufficient to explain invasion, since a particular strategy is needed for the fungus to be able to penetrate and grow within the host tissues. In the following sections we describe first the fungal strategy for invading the host tissues and secondly, the main defense mechanisms that the host could develop in order to cope with the intruder and stop the invasion process.

A. FUNGAL STRATEGY

To switch from a saprophytic to a pathogenic behaviour, *Candida* has to develop some phenotypic characteristics which allow penetration into the host organism. The propensity of the fungal cell to change its behaviour is very great and depends on its surroundings [51]; it has been shown that phenotypic switching in strains of C. albicans is associated with invasive infections [52,53]. To be successful as a pathogen *Candida* has to overcome two main obstacles: adhesion to host constituents and production of lytic enzymes. It is now well established that these two processes are associated with morphological variations. By operating a dimorphic transition [54,55] from the blastospore to the filamentous stage, C. albicans increases its adhesive properties and proteinase secretion. Many studies have shown the capacity of Candida cells, particularly the hyphal form, to bind to epithelial monoor di-saccharides by means of lectin-like surface components [56-60]. Many other fungal components were found to be involved in the linkage to epithelial cells [61-65]. In contrast, fixation to the endothelial layers requires protein-protein interactions [59,66] and may need the participation of platelets [67]. Adherence of the fungal elements to endothelial cells is followed by the internalization of fungi by an active mechanism depending on endothelial cells and is associated with production of prostaglandins [68-71].

Numerous authors have reported the fixation of the fungus to the basement membrane or to extra-cellular matrix components:

- Fixation to fibronectin is mediated by a fungal 60 kDa multimer glycoprotein [72] which presents an antigen homology with integrins α 5 β 1 and which recognizes an RGD sequence on the fibronectin molecule [73-75]. Strength of the binding increases with germ tubes and could involve interactions with different domains of fibronectin depending on the soluble or insoluble form of the molecule [76].

- Binding to laminin was shown to be mediated by a 60-68 kDa fungal mannoprotein with a quite high affinity constant [77,78]. Other linkages have been ascribed to collagen [79], entactin [80] suggesting multiple mechanisms for settlement and colonization.

The fixation capacity of *Candida* has also been demonstrated with respect to other host components :

Two kinds of complement receptors were described. A CR3-like receptor showing homologies with the CR3 of mammal cells [81,82] capable of linking to iC3b fragments and which could also be involved in the binding to epithelial cells [83,65]. A CR2-like receptor binding to C3d [84,85], the expression of which is correlated to the pathogenicity of the tested *Candida* strains [86,87]. Whereas the CR3 has homologies with the integrin family, the CR2 seems to be linked to more eclectic adhesive components capable of binding to fibrinogen, laminin, and plastic surfaces as well.

Fixation to fibrinogen is mediated by fungal mannoproteins which, by means of cross-inhibition, have been shown to be the same as those involved in binding to laminin and C3d, suggesting the idea of a multi-functionnal adhesin of *Candida* (MFA-C) [88,89] [90-92]. Taking into account the multiple roles and the ubiquity of fibrinogen throughout the body, the possibility for *Candida* to bind to this molecule with high affinity should provide different strategies for the fungus to settle and even evade from immunological surveillance with a kind of mimicry by coating its surface with soluble fibrinogen molecules belonging to the host.

Fungal cells may adhere also on prosthetic devices introduced into the host, such as catheters, cardiac valves, dental prostheses and so on. As previously described the fixation of the fungus can be mediated by the numerous host components that are quickly coated onto the foreign devices. Moreover it has also been shown, in vitro, that *Candida* can link directly to plastic surfaces by using the

MFA-C [93]. *Candida* cells which constitute the biofilm coating synthetic devices are more resistant to antifungal drugs and provide a new niche for further spread [94,95].

Following attachment to host components, *Candida* cells need to secrete enzymes to penetrate deeper in the tissues. The secretion of aspartyl proteinases of *Candida* has been extensively studied and shown to be linked to the pathogenesis of the tested strains [96-105]. Other enzymes are also involved in the pathogenesis of *Candida* [106,107].



B. HOST-DEFENSE MECHANISMS

The possibility for the fungus to settle and proliferate depends on its surroundings which is, in the present case, the host organism. Facing the fungal invasive nature, there are numerous barriers and systems which are more or less interconnected, maintaining host integrity.

At the level of the mucosal surface, *Candida* cells share the locally available nutriments with other commensal organisms. Intact skin is well protected by keratinized cells. In the normal gastro-intestinal tract, fungal cells surrounded by mucins, secretory IgA and the numerous bacteria of the flora, multiply by budding and remain more or less quiescent. But it seems that, even a slight local surface modification can trigger the pathogenic process (see iatrogenic factors). The relationship with mucins [108,109] and the status of the mucosa are major factors in determining fungal behaviour [110].

Within the blood circulation *C. albicans* has been shown to be quickly surrounded by platelets which adhere, spread over the surface of the fungus and degranulate. In mice models they are rapidly, within ten to fifteen minutes, removed from the blood stream [111,112]. This mechanism may lead to the killing of the fungal elements by small peptides secreted by activated platelets, but it may also cause the fungus to fix endothelial layers with the help of the activated platelets leading to metastatic processes [67,113,114].

Endothelial cell invasion by *C. albicans* appears to stimulate the production and extracellular secretion of

prostaglandins which are probably related to the modulation of the leukocyte response at the *Candida*-leukocyteendothelial cell interface [68]. The resistance of endothelial cells to damage provoked by *C. albicans* is increased by IFN- γ [115].

Since the surface layers are altered or crossed, an inflammatory reaction occurs. The pivotal role in the defense mechanisms against *Candida* is played by the phagocytic cells: polymorphonuclear cells (PMN) and macrophages. PMN are the first cells to reach the fighting

area within a few hours and actively cope with the invading cells. The considerable increase in candidiasis linked to neutropenia demonstrates the major role of this cell [23-25,116,117]. Anticandidal activity of granulocytopenic mice can be restored by SCF and G-CSF [118,119].

Recognition of the elements to be engulfed may be helped by opsonization mechanisms involving immunoglobulin Fc and complement receptors. Deficits in the adherence mechanisms are associated with high sensitivity to infection. PMN are very efficient because of their capability to phagocytize fungal cells and to produce lactoferrin and oxygen intermediates. Lactoferrin was demonstrated to have a high antifungal activity in vitro [120,121]; the oxydative burst and the numerous enzymes contained in the PMN granules are also responsible for the the killing of engulfed Candida cells [122,123]. More-

over, death of the PMN in the abcess area is followed by the release of a PMN-fungistatic 30 kDa protein [124]. Macrophages reach the inflammatory area secondarily. The adherence to fungal cells can also be mediated by a membrane mannose receptor [125], by means of vitronectin [126] or other receptors [127]. The fungistatic or fungicidal activity of macrophages is more often correlated to the production of nitrogen monoxyde (NO) than to that of oxygen radicals [128-130].

Migration of the phagocytic cells, activation of their metabolism and particularly the oxydative burst, increased expression of membrane receptors (FcRs, CRs) benefit largely from the local secretion of inflammatory cytokines and particularly IFNs and TNFs [131-134] which in turn, can be modulated by *Candida* components [135-137]. Other cytokines are also involved in these mechanisms: IL-2 [138], IL-8 [139], GM-CSF [140].

The local level of cytokines is largely dependent on the antigen specific recognition operated by T lymphocytes. Two different subsets of T helper cells were described (Th1 and Th2) and shown to play antagonistic roles. Th1 is associated with IFN- γ , IL-2 production and Th2 with IL-4 and IL-10 mainly. Th1 corresponds to what has long been designated as cellular immunity while Th2 controls the humoral response. By means of experimental murine models, it was demonstrated that protection against mucocutaneous candidiasis is associated with a Th1 response and in contrast the disease was linked to a Th2 reactivity [141]. Activation of Th1 rather than Th2 may depend on various factors, such as the antigen presenting cell, the HLA II molecules involved, the presence

of particular cytokines and also the steric configuration of the antigen. Artificial protein conjugated mannan can direct a Th1 response when obtained in oxidizing conditions and a Th2 response when conjugated under reducing conditions [142]. These results present considerable interest regarding the possibility of driving the immune reactivity toward an efficient response [143]. However the correlation between the local immune response and the general immune status is difficult to establish and results remain controversial [144,145].

The protective role of humoral antibodies in the resolution of systemic candidiasis is not well established. Despite results showing the increase of anti-enolase antibodies [146] and anti-C. albicans heat shock protein (HSP) 90 [147] in patients recovering from systemic candidiasis, there is no clear correlation between humoral immunity and resistance or susceptibility to infection with C. albicans [148]. Nevertheless, at the mucosal level, IgA secretion could be involved in the local protection [149-151].

HSP also called "chaperones" are highly conserved throughout species evolution [152] suggesting their major role in cell biology. Candidal HSP 90, which appears to be a major antigen in Candida infections, shares common epitopes with human HSP, leading to possible interferences between self and non-self reactivity [147]. Other HSP were described in *Candida* [153,154]. HSP are mainly involved in repair and refolding of denatured proteins and could be also involved in antigen processing. They are supposed to intervene in the antigen recognition of the T (CD4-/CD8-) $\gamma\delta$ cells. Taking into account this property, the location of T $\gamma\delta$ cells in the subepithelial area and the fact that they are able to produce large amounts of IFN- γ , it could be hypothetized that they play an important role in the modulation of local inflammatory reaction [155,156] and deserve to be further studied in candidiasis.

CYTOKINE INTERPLAY



CONCLUSION

In the context of immunodeficient hosts, Candida may behave as a true pathogen and proliferate until confronted by an efficient defense mechanism. The balance is dynamic with, on one hand the multiple barriers of host immunity, and on the other the burden of the invading cells. Recent progress in the understanding of the cellular and molecular mechanisms involved in mucocutaneous candidiasis raises hopes of possible control of the immune response. Considering that about one-third of immunosuppressed patients affected by Candida septicemia die, and taking into account the fact that iatrogenic factors lead to the selection of more pathogenic strains and to the emergence of species resistant to fungal therapies, the best medical attitude is obviously to survey and prevent a possible extension of Candida cells of endogenous or exogenous origin.

References

- 1. Beck-Sague CM, Jarvis WR. Secular trends in the epidemiology of nosocomial fungal infections in the United Sates,1980-1990. J Infect Dis 1993;167:1247-1251.
- 2. Jarvis WR. Epidemiology of nosocomial fungal infections, with emphasis on Candida species. Clin Infect Dis 1995;20:1526-1530
- Fisherhoch SP, Hutwagner L. Opportunistic candidiasis: An epidemic of the 1980s. Clin Infect Dis 1995;21:897-904.
- Muller J. The epidemiology of deep-seated, domestic mycoses. Mycoses 1994;37 Suppl. 2):1-7.
- Koll B, Brown A. The changing epidemiology of infections at cancer hospitals. Clin Infect Dis 1993;17(Suppl. 2):322-328.
- 6. Wingard JR. Importance of Candida species other than C. albicans as pathogens in oncology patients. Clin Infect Dis 1995;

20:115-125.

- Pfaller MA. Infection control: opportunistic fungal infections-the increasing importance of *Candida* sp. Infect Control Hosp Epidemiol 1989;9:408-416.
- Pfaller MA. Epidemiology and control of fun-gal infections. Clin Infect Dis 1994;19 (Suppl. 1):S8-S13.
- 9. Meunier F, Aoun M, Bitar N. Candidemia in immunocompromised patients. Clin Infect Dis 1992;14 (Suppl. 1):S120-S125. 10. Wingard JR. Infections due to resistant
- Candida species in patients with cancer who are receiving chemotherapy. Clin Infect Dis 1994;19 (Suppl. 1):S49-S53.
- 11.Boken D, Swindells S, et al. Fluconazoleresistant Candida albicans. Clin Infect Dis 1993:17:1018-1021
- 12.Hazen KC. New and emerging yeast patho-

- gens. Clin Microbiol Rev 1995;8:462. 13.Odds FC. Epidemiological shifts in opportu-nistic and nosocomial *Candida* infections: Mycological aspects. Int J Antimicrob Agents 1996:6:141-144.
- 14. Pfaller MA. Nosocomial candidiasis: Emerging species, reservoirs, and modes of transmission. Clin Infect Dis 1996;22 (Suppl. 2):S89-S94.
- 15. Romano F, Ribera G, Giuliano M. A study of a hospital cluster of systemic candidosis using DNA typing methods. Epidemiol Infect 1994;112:393-398.
- Vazquez JA, Sanchez V, Dmuchowski C, Dembry LM, Sobel JD, Zervos MJ. Nosocomial Acquisition of Candida albicans an epidemiologic study. J Infect Dis 1993; 168:195-201
- 17. Rangel-Frausto M, Martin M, Saiman L, et

al. High-prevalence of Candida species on hands care workers in surgical and neonatal intensive care units: a multicenter study. 1994: 34th ICAAC: Poster #105.

- Spinillo A, Capuzzo E, Nicola S, Baltaro F, Ferrari A, Monaco A. The impact of oral con-
- Ferrari A, Monaco A. The Impact of oral contraception on vulvovaginal candidiasis. Contraception 1995;51:293-297.
 19. Nohmi T, Abe S, Dobashi K, Tansho S, Yamaguchi H. Suppression of anti-*Candida* activity of murine neutrophils by progesterone in vitro: A possible mechanism in pregnant women's vulnerability to vaginal. nant women's vulnerability to vaginal candidiasis. Microbiol Immunol 1995;39:405-409.
- Zhao X, Malloy PJ, Ardies CM, Feldman D. Oestrogen-binding protein in *Candida albi-cans*: Antibody development and cellular localization by electron immunocytoche-mistry. Microbiol-UK 1995;141(Part
- 2685-2692.
 Caticha O, Odell WD. Characterization and Purification of the chorionic gonadotropin-like protein binding site in *Candida albicans* Endocrin Res 1994;20:1-19.
- 22. Reszel E, Mishra S, Mishra J, Mishra A Pierson D. Stress, immunity and mucocuta-neous candidiasis. J Osteopath Med
- neous candidiasis. J Osteopatri vieu 1993;7:26-28.
 23. Martino P, Girmenia C, Micozzi A, *et al.* Fungemia in patients with leukemia. Am J Med Sci 1993;306:225-232.
 24. Swerdloff JN, Filler SG, Edwards JE. Severe candidal infections in neutropenic opticate. Clin olfact Dis 1903;17 (Suppl. 2
- patients. Clin Infect Dis 1993;17 (Suppl. 2): S457-S467
- 25. Jensen J, Warner T, Balish E. The role of phagocytic cells in resistance to disseminated candidiasis in granulocytopenic mice. J Infect Dis 1994;170:900-905.
- Kirkpatrick CH. Chronic mucocutaneous candidiasis. J Am Acad Dermatol 1994; 31
- (Suppl. 3, P2):S14-S17.
 27. Segal E, Joseflev A. Induction of candidal vaginitis in diabetic mice and attempts to prevent the infection. J Med Vet Mycol 1995; 33.1-8
- 28. Kennedy MJ, Volz PA. Effect of various antibiotics on gastrointestinal colonization and dissemination by *Candida albicans*. Sabouraudia: J Med Vet Mycol 1985;23:265-
- 29. Vanwyk CW, Vanderbijl P, Stander I. Enhanced resistance to Candida albicans in mice during tetracycline administration. South Afr J Sci 1995;91:140-141. 30. Levine J, Dykoski RK, Janoff EN. *Candida*-
- associated diarrhea: A syndrome in search of credibility. Clin Infect Dis 1995;21:881-886.
- 31. Bernhardt H, Wellmer A, Zimmermann K, Knoke N. Growth of *Candida albicans* in normal and altered faecal flora in the model of continuous flow culture. Mycoses 1995;38: 265-270
- 32. Braun PC. Surface hydrophobicity enhances corticosterone incorporation in *Candida albicans*. Infect Immun 1994;62:4087-4090.
- 33. Cavaillon JM. Contribution of cytokines to inflammatory processes. Pathol Biol 1993; 1:799-811
- 34. Nohmi T, Abe S, Tansho S, Yamaguchi H. Suppression of anti-Candida activity of muri-ne and human neutrophils by glucocorti-coids. Microbiol Immunol 1994;38:977-982.
 Heidenreich S, Kubis T, Schmidt M, Fegeler W. Glucocorticoid-induced alterations of version of the proceeding of the second second term of the second second
- monocyte defense mechanisms against Candida albicans. Cell Immunol 1994;157:320-327
- Deslauriers N, Coulombe C, Carre B, Goulet JP. Topical application of a corticosteroid destabilizes the host-parasite relationship in an experimental model of the oral carrier state of Candida albicans. FEMS Immunol
- Med Microbiol 1995;11:45-55. 37. Botas CM, Kurlat I, Young SM, Sola A. Disseminated candidal infections and intravenous hydrocortisone in preterm infants Pediatrics 1995;95:883-887.
- 38. Denning DW. Epidemiology and pathogenesis of systemic fungal infections in the immu-nocompromised host. J Antimicrob
- Chemother 1991;28:1-16.39. Chanock S. Evolving risk factors for infectious complications of cancer therapy. Hematol Oncol Clin North Amer 1993; 7: 771-793
- 40. Ueta E, Osaki T, Yoneda K, Yamamoto T,

Umazume M. Influence of inductive chemoradiotherapy on salivary polymorphonuclear leukocyte (SPMN) functions in oral cancer. J Oral Pathol Med 1994;23:418-422.

- 41. Mudad R, Vredenburgh J, Paulson EK, et al. A radiologic syndrome after high dose che-A facility syndrome after might dose che-motherapy and autologous bone marrow transplantation, with clinical and pathologic features of systemic candidiasis. Cancer 1994; 74:1360-1366.
 42. Sandovskylosica H, Barrnea L, Segal E. Fatal systemic candidiasis of gastrointestinal origin an experimental media in piec com
- origin an experimental model in mice com-promised by anti-cancer treatment. J Med
- Vet Mycol 1992;30:219-231.
 43. Larner AJ, Hamilton MIR. Infective complications of therapeutic gastric acid inhibition. Alimentar Pharmacol Ther 1994;8:579-584.
 44. Deptide A. Nijeiroschi L. Jetterane On. Each
- Rantala A, Niinikoski J, Lehtonen OP. Early Candida isolations in febrile patients after abdominal surgery. Scand J Infect Dis 1993; 25.479-485
- 45. Pappo I, Polacheck I, Zmora O, Feigin E, Freund HR. Altered gut barrierfFunction to Candida during parenteral nutrition. Nutrition 1994;10:151-154.
- Lee J, Navarro E, et al.
 Vascular catheter-associated fungemia in patients with cancer: analysis of 155 episo-des. Clin Infect Dis 1992;14:875-883.
 Raad I, Bodey G. Infectious complications of interest Distance of the sector Distance of Dista
- indwelling vascular catheters. Clin Infect Dis 1992:15:197-210.
- 48. Stickler DJ, Mclean RJC. Biomaterials associated infections: The scale of the problem. Cell Mater 1995;5:167-182.
- 49. Meyers J. Fungal infections in bone marrow transplant patients. Semin Oncol 1990;17: 10-13
- 50. Morrison VA, Haake RJ, Weisdorf DJ. Noncandida fungal infections after bone marrow transplantation: Risk factors and outcome. Am J Med 1994;96:497-503
- 51. Mccullough MJ, Ross BC, Dwyer BD, Reade PC. Genotype and phenotype of oral *Candida albicans* from patients infected with the human immunodeficiency virus. Microbiol-UK 1994;140:1195-1202. 52. Jones S, White G, Hunter PR. Increased
- phenotypic switching in strains of *Candida* albicans associated with invasive infections.
- J Clin Microbiol 1994;32:2869-2870. 53. De Bernardis F, Adriani D, Lorenzini R, Pontieri E, Carruba G, Cassone A. Filamentous growth and elevated vaginopat-hic potential of a nongerminative variant of *Candida albicans* expressing low virulence in systemic infection. Infect Immun 1993;61: 1500-1508.
- 54. Tronchin G, Bouchara JP, Robert R Dynamic changes of the cell wall surface of *Candida albicans* associated with germina-tion and adherence. Eur J Cell Biol 1989;50: 285-290
- 55. Gow NAR, Perera THS, Sherwoodhigham J, Gooday GW, Gregory DW, Marshall D. Investigation of touch sensitive responses by hyphae of the human pathogenic fungus Candida albicans. Scan Micros 1994;8:705-710.
- 56. Critchley IA, Douglas LJ. Role of glycosides as epithelial cell receptors for *Candida albicans*. J Gen Microbiol 1987;133:637-643.
- 57. Douglas LJ. Adhesion of Candida albicans
- to host surfaces. *Candida* and Candidamycosis 1991;50:43-47. 58. Tosh FD, Douglas LJ. Characterization of a
- fucoside-binding adhesin of *Candida albi-cans*. Infect Immun 1992;60(11):4734-4739.
 Calderone R, Wadsworth E. Adherence molecules of *Candida albicans*. J Microbiol Methods 1993;18:197-211.
- Cameron BJ, Douglas LJ. Blood group gly-colipids as epithelial cell receptors for Candida albicans. Infect Immun 1996;64 891-896. 61. Li RK. Cutler JE. Chemical Definition of an
- epitope/adhesin molecule on Candida albi-
- cans. J Biol Chem 1993;268:18293-18299.
 62. Kanbe T, Cutler JE. Evidence for adhesin activity in the acid-stable moiety of the phosphomannoprotein cell wall complex of Candida albicans. Infect Immun 1994;62: 1662-1668.
- 63. Yu L, Lee KK, Sheth HB, et al. Fimbria-mediated adherence of Candida albicans to glycosphingolipid receptors on human buccal epithelial cells. Infect Immun 1994;62:2843-2848

- 64. Lehrer N, Segal E, Lis H, Gov I. Effect of Candida albicans cell wall components on the adhesion of the fungus to human and murine vaginal mucosa. Mycopathologia 1988;102:115-121.
 Bendel CM, Stsauver J, Carlson S, Hostetter MK. Epithelial adhesion in yeast species: Correlation with surface avpression
- species: Correlation with surface expression of the integrin analog. J Infect Dis 1995;171: 1660-1663
- Edwards JE, Mayer CL, Filler SG, Wadsworth E, Calderone RA. Cell extracts of Candida albicans block adherence of the organisms to endothelial cells. Infect Immun 1992;60:3087-3091
- Klotz S, Harrison J, Misra R. Aggregated platelets enhance adherence of *Candida* yeasts to endothelium. J Infect Dis 1989:160:669-677.
- 68. Filler SG, Ibe BO, Ibrahim AS, Ghannoum MA, Raj JU, Edwards JE. Mechanisms by which Candida albicans induces endothelial cell prostaglandin synthesis. Infect Immun
- 1994;62:1064-1069. 69. Filler SG, Swerdloff JN, Hobbs C, Luckett PM. Penetration and damage of endothelial cells by Candida albicans. Infect Immun 1995;63:976-983.
- Calderone R, Diamond R, Senet JM, Warmington J, Filler S, Edwards JE. Host cell-fungal cell interactions. J Med Vet Mycol 1994;32:151-168.
- Klotz SA. Adherence of *Candida albicans* to endothelial cells is inhibited by prostaglandin I-2. Infect Immun 1994;62:1497-1500.
 Klotz SA, Hein RC, Smith RL, Rouse JB. The fibronectin adhesin of *Candida albicans*.
- Infect Immun 1994;62:4679-4681.
 73. Penn C, Klotz SA. Binding of plasma fibro-nectin to *Candida albicans* occurs through the cell binding domain. Microb Pathogen 1994:17:387-393.
- 74. Santoni G, Gismondi A, Liu JH, et al. Candida albicans expresses a fibronectin receptor antigenically related to alpha 5 beta 1 integrin. Microbiol-UK 1994;140:2971 2979
- Zara.
 Santoni G, Birarelli P, Hong LJ, Gamero A, Djeu JY, Piccoli M. An alpha 5 beta 1 -like integrin receptor mediates the binding of less pathogenic *Candida* species to fibronectin. J
- Med Microbiol 1995;43:360-367.
 76. Negre E, Vogel T, Levanon A, Guy R, Walsh TJ, Roberts DD. The collagen binding domain of fibronectin contains a high affinity binding site for *Candida albicans*. J Biol Chem 1994;269:22039-22045.
- 77. Bouchara J, Tronchin G, Annaix V, Robert R, Senet J. Laminin receptors on *Candida*
- R, Senet J. Laminin receptors on *Canada* albicans germ tubes. Infect Immun 1990;58:48-54.
 78. Lopez-Ribot JL, Casanova M, Monteagudo C, Sepulveda P, Martinez JP. Evidence for the presence of a high-affinity laminin recep-tor-like molecule on the surface of *Canadia* albicant and a surface of *Canadia* albicans yeast cells. Infect Immun 1994;62: 742-746
- 79. Klotz SA, Smith RL. Gelatin fragments block adherence of Candida albicans to extracellular matrix proteins. Microbiol-UK 1995;141: 2681-268
- 80. Lopez-Ribot JL, Chaffin WL. Binding of the extracellular matrix component entactin to Candida albicans. Infect Immun 1994;62: 4564-4571
- 4304-4371.
 81. Gilmore B, Retsinas E, Lorenz J, Hostetter M. An iC3b receptor on *Candida albicans* :
- structure, function and correlates for patho-genicity. J Infect Dis 1988;157:38-46. . Alaei S, Larcher C, Ebenbichler C, Prodinger WM, Janatova J, Dierich MP. Isolation and biochemical characterization of 82. the iC3b receptor of *Candida albicans*. Infect Immun 1993;61:1395-1399.
- Hostetter MK. Adhesins and ligands involved in the interaction of *Candida* spp with epithelial and endothelial surfaces. Clin Microbiol Rev 1994;7:29-42.
- Calderone RA, Linehan L, Wadsworth E, Sandberg AL. Identification of C3d receptors on *Candida albicans*. Infect Immun 1988;56: 252-258
- 85. Wadsworth E, Prasad SC, Calderone R. Analysis of mannoproteins from blastoconidia and hyphae of *Candida albicans* with a common epitope recognized by anti-complement receptor type-2 antibodies. Infect Immun 1993;61:4675-4681.
- 86. Fukayama M, Wadsworth E, Calderone R.

Expression of the C3d-binding protein (CR2) from *Candida albicans* during experimental candidiasis as measured by lymphoblasto-genesis. Infect Immun 1992;60:8-12.

- Franzke S, Calderone RA, Schaller K. Isolation of avirulent clones of *Candida albi* cans with reduced ability to recognize the CR2 ligand C3d. Infect Immun 1993;61:
- 2662-2669.
 88. Bouali A, Robert R, Tronchin G, Senet JM. Characterization of binding of human fibrino-gen to the surface of germ-tubes and mycelium of *Candida albicans*. J Gen Microbiol 1987;133:545-551.
- Annaix V, Bouchara J, Tronchin G, Senet J, Robert R. Structures involved in the binding of human fibrinogen to *Candida albicans* germ tubes. FEMS Microbiol Immunol 1990; 64:147-154.
- 64:147-154.
 90. Robert R, Mahaza C, Marot-Leblond A, Tronchin G, Senet JM. Binding of mouse fibrinogen to *Candida albicans* in vivo. FEMS Microbiol Lett 1991;78:301-304.
- Casanova M, Lopez-Ribot JL, Monteagudo C, Llombartbosch A, Sentandreu R, Martinez JP. Identification of a 58-kilodalton cell surface fibrinogen-binding mannoprotein from Candida albicans. Infect Immun 1992; 60:4221-4229.
- Martinez JP, Lopez-Ribot JL, Chaffin WL Heterogeneous surface distribution of the fibrinogen-binding protein on Candida albi-
- cans. Infect Immun 1994;62:709-712.
 93. Tronchin G, Bouchara J, Robert R, Senet J. Adherence of *Candida albicans* germ tubes to plastic: ultrastructural and molecular studies of fibrilar adhesins. Infect Immun
- 1988;56:1987-1993.
 94. Hawser SP, Douglas LJ. Biofilm formation by *Candida* species on the surface of catheter materials in vitro. Infect Immun 1994;62: 915-921
- Hawser SP, Douglas LJ. Resistance of Candida albicans biofilms to antifungal agents in vitro. Antimicrob Agents
- agents *II* virio. Antificiolo Agents Chemother 1995;39:2128-2131.
 96. Borg M, Ruchel R. Expression of extrace-llular acid proteinase by proteolytic *Candida* spp. during experimental infection of oral mucosa. Infect Immun 1978;56:626-631.
 97. Rüchel R, De Bernardis F, Ray T, Sullivan B, Cole C, Condida acid proteinaces. IMdd
- P, Cole G. *Candida* acid proteinases. J Med Vet Mycol 1992;30 (Suppl.1):123-132.
 98. White TC, Agabian N. *Candida albicans*
- White TC, Agabian N. Candida albicans secreted aspartyl proteinases: Isoenzyme pattern is determined by cell type, and levels are determined by environmental fac-tors. J Bacteriol 1995;177:5215-5221.
 De Bernardis F, Cassone A, Sturtevant J, Calderone R. Expression of *Candida albi-cans* SAP1 and SAP2 in experimental vagi-nitis. Infect Immun 1995;63:1887-1892.
 De Bernardis F, Chiani P, Ciccozzi M, et al. Elevated aspartic proteinase secretion
- al. Elevated aspartic proteinase secretion and experimental pathogenicity of Candida albicans isolates from oral cavities of subjects infected with human immunodeficiency virus. Infect Immun 1996;64:466-471. 101. Hoegl L, Ollert M, Korting HC. The role of
- Candida albicans secreted aspartic proteinase in the development of candidoses. J
- Mol Med 1996;74:135-142. 102. Hube B, Monod M, Schofield DA, Brown AJP, Gow NAR. Expression of seven members of the gene family encoding secretory aspartyl proteinases in *Candida albicans*. Mol Microbiol 1994;14:87-99.
 103. Kaminishi H, Miyaguchi H, Tamaki T, *et al*. Degradation of humoral host defense by
- Candida albicans proteinase. Infect Immun 1995;63:984-988
- 1995;63:984-988.
 104. Louie A, Dixon DM, Elmaghrabi EA, Burnett JW, Baltch AL, Smith RP. Relation-ship between *Candida albicans* epidermoly-tic proteinase activity and virulence in mice. J Med Vet Mycol 1994;32:59-64.
 105. Ollert MW, Wende C, Gorlich M, et al. Increased expression of *Candida albicans* secretory profession of *Candida albicans*
- secretory proteinase, a putative virulence factor, in isolates from human immunodeficiency virus-positive patients. J Clin
- Microbiol 1995;33:2543-2549.
 106. Nikai T, Okumura Y, Hasegawa Y, Uchiya K, Kamiya K, Sugihara H. Isolation and characterization of fibrinogenase from *Candida albicans* NH-1. Int J Biochem 1993;25:1815-1822
- 107. Ibrahim AS, Mirbod F, Filler SG, et al. Evidence implicating phospholipase as a

virulence factor of Candida albicans. Infect

- Immun 1995;63:1993-1998. 108. Hoffman MP, Haidaris CG. Analysis of *Candida albicans* adhesion to salivary
- mucin. Infect Immun 1993;61:1940-1949. 109. Hoffman MP, Haidaris CG. Identification and characterization of a Candida albicans binding proteoglycan secreted from rat submandibular salivary glands. Infect Immun 1994;62:828-836.
- 110. Rindum JL, Stenderup A, Holmstrup P. Identification of Candida albicans types related to healthy and pathological oral mucosa. J Oral Pathol Med 1994;23:406-412.
- 1. Mahaza C, Robert R, Miegeville M, Tronchin G, Senet JM. *Candida albicans* -platelet interaction evidence for *in vivo* and in vitro, cell to cell attachment. In: Tümbay E (Ed.) *Candida* and Candidamycosis. New York, Plenum Press, 1991: 131-135.
- Mahaza C, Robert R, Senet JM. Candida albicans-platelet interaction: molecules involved in the adherence. In: Tümbay E
- (Ed.) Candida and Candidamycosis. New York, Plenum Press, 1991: 137-141.
 113. Yeaman MR, Ibrahim AS, Edwards JE, Bayer AS, Ghannoum MA. Thrombin-indu-ced rabbit platelet microbicidal protein is fungicidal in vitro. Antimicrob Agents
- Chemother 1993;37:546-553. 114. Yeaman MR, Soldan SS, Ghannoum MA, Edwards JE, Filler SG, Bayer AS. Resistance to platelet microbicidal protein results in increased severity of experimental *Candida albicans* endocarditis. Infect Immun
- 1996;64:1379-1384. 115. Ibrahim AS, Filler SG, Ghannoum MA, Edwards JE. Interferon-gamma protects endothelial cells from damage by *Candida albicans*. J Infect Dis 1993;167:1467-1470. 116. Martino P, Girmenia C, Venditti M. *Candida* colonization and systemic infection
- Candida colonization and systemic infection in neutropenic patients: A retrospective study. Cancer 1989;64:2030-2034.
 117. Sohnle PG, Hahn BL, Wagner DK. Arrays of *Candida albicans* pseudohyphae that pro-to arbitrary provide the providence of the provide
- tect the organisms from neutrophil fungicidal mechanisms in experimental infections of
- mice. J Med Vet Mycol 1994;32:21-30.
 118. Steinshamn S, Bergh K, Waage A. Effects of stem cell factor and granulocyte colonystimulating factor on granulocyte ecovery and *Candida albicans*-infection in granulocy-topenic mice. J Infect Dis 1993; 168:1444-1448
- 119. Hamood M, Bluche PF, Devroey C, Corazza F, Bujan W, Fondu P. Effects of recombinant human granulocyte-colony sti-mulating factor on neutropenic mice infected with *Candida albicans*: Acceleration of recovery from neutropenia and potentiation of anti-C. albicans resistance. Mycoses 1994; 37:93-99.
- Bellamy W, Wakabayashi H, Takase M, Kawase K, Shimamura S, Tomita M. Killing of *Candida albicans* by lactoferricin-B, a potent antimicrobial peptide derived from the N-terminal region of bovine lactoferrin. Med
- Microbiol Immunol 1993;182:97-105. 121. Nikawa H, Samaranayake LP, Tenovuo J, Pang KM, Hamada T. The fungicidal effect of human lactoferrin on Candida albicans and Candida krusei. Arch Oral Biol 1993;38: 1057-1063
- 122. Diamond R, Clark R, Haudenschild C. Damage to Candida albicans hyphae and pseudohyphae by the myeloperoxidase sys tem and oxydative products of neutrophil metabolism in vitro. J Clin Invest 1980;66: 908-917
- 123. Stein DK, Malawista SE, Vanblaricom G, Wysong D, Diamond RD. Cytoplasts gene-rate oxidants but require added neutrophil granule constituents for fungicidal activity against Candida albicans hyphae. J Infect
- Dis 1995;172:511-520. 124. McNamara M, Weissner J, Collins-Lech C, Hahn B, Sohnle P. Neutrophil death as a defence mechanism against Candida albi-cans infections. Lancet 1988;2:1163-1165
- 125. Kitz DJ, Stahl PD, Little JR. The Effect of a mannose binding protein on macrophage interactions with Candida albicans. Cell Mol Biol 1992;38:407-412.
- 126. Limper AH, Standing JE. Vitronectin inte-racts with Candida albicans and augments organism attachment to the NR8383 macrophage cell line. Immunol Lett 1994;42:139-144.

- 127. Szabo I, Guan LM, Rogers TJ. Modulation of macrophage phagocytic activity by cell wall components of *Candida albicans*. Cell Immunol 1995;164:182-188.
- Blasi E, Pitzura L, Puliti M, *et al.* Differential susceptibility of yeast and hyphal forms of *Candida albicans* to macrophagederived nitrogen-containing compounds. Infect Immun 1995;63:1806-1809.
- Milect minint 1935,053,1000-1005.
 129. Rementeria A, Garcia Tobalina R, Sevilla MJ. Nitric oxide-dependent killing of *Candida* albicans by murine peritoneal cells during an experimental infection. FEMS Immunol Med Microbiol 1995;11:157-162.
- 130. Vazqueztorres A, Jonescarson J, Balish E. Vazqueziones A, Jonescarson J, Ballshi F. Nitric oxide production does not directly increase macrophage candidacidal activity. Infect Immun 1995;35:1142-1144.
 Watanabe K, Kagaya K, Yamada T, Fukazawa Y. Mechanism for candidacidal
- activity in macrophages activated by recombinant gamma-interferon. Infect Immun 1991;59:521-528.
- 132. Steinshamn S, Waage A. Tumor necrosis factor and IL-6 in *Candida albicans* infection Inormal and granulopenic mice. Infect Immun 1992;60:4003-4008.
 133. Diamond R, Lyman C, Wysong D. Disparate effects of interferon-gamma and
- TNF-alpha on early neutrophil respiratory burst and fungicidal responses to Candida albicans hyphae in vitro. J Clin Invest 1991; 87.711-720
- 134. Stevenhagen A, Vanfurth R. Interferongamma activates the oxidative killing of *Candida albicans* by human granulocytes. Clin Exp Immunol 1993;91:170-175.
- 135. Garner RE, Rubanowice K, Sawyer RT, Hudson JA. Secretion of TNF-alpha by alve-olar macrophages in response to *Candida albicans* mannan. J Leukoc Biol 1994;55: 161-168
- 136. Jouault T, Lepage G, Bernigaud A, et al. beta-1,2-linked oligomannosides from Candida albicans act as signals for tumor
- Candida albicans act as signals for tumor necrosis factor alpha production. Infect Immun 1995;63:2378-2381.
 137. Castro M, Bjoraker JA, Rohrbach MS, Limper AH. Candida albicans induces the release of inflammatory mediators from human peripheral blood monocytes. Information 1996;20:107.122
- Inflammation 1996;20:107-122. 138.Djeu JY, Liu JH, Wei S, *et al.* Function associated with IL-2 receptor-beta on human neutrophils - mechanism of activation of anti-fungal activity against *Candida albicans* by IL-2. J Immunol 1993;150:960-970.
- Djeu J, Matsushima K, Oppenheim J, Shiotski K, Blanchard D. Functionnal activation of human neutrophils by recombinant monocyte-derived neutrophil chemotactic factor/IL-8. J Immunol 1990;144:2205-2210. 140. Wang M, Friedman H, Djeu J.
- Enhancement of human monocyte function against *Candida albicans* by the colony-sti-mulating factors (CSF): IL-3, granulocyte macrophage CSF, and macrophage CSF. J Immunol 1989;143:671-677.
- 141. Romani L, Howard DH. Mechanisms of resistance to fungal infections. Curr Opin Immunol 1995;7:517-523.
- 142. Apostolopoulos V, Pietersz GA, Loveland BE, Sandrin MS, Mckenzie IFC. Oxidative / reductive conjugation of mannan to antigen selects for T-1 or T-2 immune responses. Proc Natl Acad Sci USA 1995; 92: 10128-10132
- 143. Mencacci A, Cenci E, Spaccapelo R, Tonnetti L, Romani L. Rationale for cytokine
- and anti-cytokine therapy of *Candida albi-cans* infection. J Mycol Med 1995;5:25-30.
 Mencacci A, Torosantucci A, Spaccapelo R, Romani L, Bistoni F, Cassone A. A mannoprotein constituent of *Candida albicans* that elicits different levels of delayed-type hypersensitivity, cytokine production, and anticandidal protection in mice. Infect Immun 1994;62:5353-5360.
- 145. Garner RE, Domer JE. Lack of Effect of Candida albicans mannan on development of protective immune responses in experimental murine candidiasis. Infect Immun 1994;62:738-741.
- 1994,02.736-741.
 146. Franklyn K, Warmington J, Ott A, Ashman R. An immunodominant antigen of *Candida albicans* shows homology to the enzyme enolase. Immunol Cell Biol 1990;68:173-178.
- 147. Matthews RC. Candida albicans HSP-90 link between protective and auto Immunity. J

12

- Med Microbiol 1992;36:367-370.
 148. Costantino PJ, Gare NF, Warmington JR. Humoral immune responses to systemic *Candida albicans* infection in inbred mouse strains. Immunol Cell Biol 1995;73:125-133.
 149. Vudhicharmong K, Walker DM, Ryley HC. The effect of secretory immunoglobulin A on the *in vitro* adherence of the yeast *Candida albicans* to human oral epithelial cells. Arch Oral Biol 1982;27:617-621.
 150. Coogan MM, Sweet SP, Challacombe SJ. Immunoglobulin A (IgA), IgA1, and IgA2 Antibodies to *Candida albicans* in whole and parotid saliva in human immunodeficiency virus infection and AIDS. Infect Immun 1994; virus infection and AIDS. Infect Immun 1994; 62:892-896.
- 151. Umazume M, Ueta E, Osaki T. Reduced inhibition of Candida albicans adhesion by
- Initiation for patients receiving oral cancer therapy. J Clin Microbiol 1995;33:432-439.
 Swoboda RK, Bertram G, Budge S, Gooday GW, Gow NAR, Brown AJP.
 Structure and regulation of the HSP90 gene from the aetherapic function. Condition of the from the pathogenic fungus Candida albi-

cans. Infect Immun 1995;63:4506-4514.

- 153. Lavalle R, Bromuro C, Ranucci L, Muller HM, Crisanti A, Cassone A. Molecular clo-ning and expression of a 70-kilodalton heat shock protein of *Candida albicans*. Infect Immun 1995;63:4039-4045.
- 154. Maresca B, Kobayashi GS. Hsp70 in parasites: As an inducible protective protein and as an antigen. Experientia 1994;50:1067-1074.
- 155. Chakir J, Cote L, Coulombe C, Deslauriers
 N. Differential pattern of infection and immu-
- N. Differential patients of intervential and initial or an error of the patient of the oxide production enhances resistance to mucosal candidiasis. Nature Med 1995;1:552-557.