

A new caffeic acid minimal synthetic medium for the rapid identification of *Cryptococcus neoformans* isolates

Valerio Vidotto¹, Shigeji Aoki², José Pontón³, Guillermo Quindós³, Cristiane Yumi Koga-Ito¹ & Agostino Pugliese¹

¹Dipartimento Discipline Medico-Chirurgiche, Università di Torino, Sezione Malattie Infettive, Torino, Italy; ²Molecular Biology Laboratory, Advanced Research Center, Nippon Dental University, Niigata, Japan; ³Departamento de Inmunología, Microbiología y Parasitología, Facultad de Medicina y Odontología, Universidad del País Vasco, España

Summary	Melanin production is one of the most important criteria for rapid identification of <i>Cryptococcus neoformans</i> . Most of the media described in the literature for identifying <i>C. neoformans</i> are very complex; they contain many organic or inorganic compounds and are difficult to prepare and store. The new minimal synthetic caffeic acid medium described in this paper is simpler to prepare, convenient and constitutes an interesting new medium for the rapid identification of <i>C. neoformans</i> isolates.				
Key words	Cryptococcus neoformans, Caffeic acid medium, Phenoloxidase, Identification				
	Un nuevo medio sintético mínimo para la identificación rápida de aislamientos de <i>Cryptococcus neoformans</i>				
Resumen	La producción de melanina es uno de los criterios más importantes para la identificación rápida de <i>Cryptococcus neoformans</i> . La mayoría de los medios de cultivo descritos para la identificación de <i>C. neoformans</i> son muy complejos: contienen muchos compuestos orgánicos e inorgánicos y son difíciles de preparar y almacenar. El nuevo medio mínimo sintético con ácido caféico, descrito en este trabajo, es más sencillo de preparar y constituye un nuevo medio apropiado para la identificación rápida de <i>C. neoformans</i> .				
Palabras clave	<i>Cryptococcus neoformans</i> , Medio con ácido caféico, Fenoloxidasa, Identificación				

Mycoses are a growing medical problem requiring a prompt diagnosis to facilitate an early antifungal treatment. *Cryptococcus neoformans* has risen to a worldwide highly recognizable major opportunistic pathogen with deadly consequences in immunocompromised patients, particu-

Address for correspondence: Dr. Valerio Vidotto Dipartimento Discipline Medico-Chirurgiche Università di Torino, Sezione Malattie infettive Corso Svizzera 164 10.149 Torino, Italy Tel.: +39 011 439 3860 Fax: +39 011 439 3882 E-mail: valerio.vidotto@libero.it

Aceptado para publicación el 2 de julio de 2004

©2004 Revista Iberoamericana de Micología Apdo. 699, E-48080 Bilbao (Spain) 1130-1406/01/10.00 Euros larly in those with AIDS. Cryptococcosis, in the pre-high activity anti-retroviral therapy era, became a major cause of disease and death among patients with AIDS and was the commonest life-threatening mycosis in these patients. Drug users, homosexual men, polytransfunded patients, hemophiliacs, and immunocompromised hosts, among others, are at high risk of cryptococcosis [3,9,15].

Melanin production by phenoloxidase activity is a distinctive and characteristic property of C. neoformans isolates [2,4,6]. The ability to produce these melanin pigments is one of the most used criteria for the identification of C. neoformans clinical and environmental isolates and for the evaluation of Cryptococcus virulence [13,17]. Melanin production is usually tested in a proper agar medium containing a precursor of melanin. For this purpose, agar media containing DOPA, caffeic acid, birdseed or sunflower extracts have been reported so far [1,5-8,10-12,14,16,18,19]. Vidotto et al. [20] have developed a vitamin-free minimal synthetic medium for C. neoformans. In the present paper, we report the comparison of melanin production by clinical and environmental isolates of C. neoformans and other medically important yeasts on a new minimal synthetic caffeic acid medium (MSCAM).

Materials and methods

The original caffeic acid agar medium (CAAM) -composition per liter: glucose 5 g; ammonium sulphate 5 g; yeast extract 2 g; potassium phosphate 0.8 g; magnesium sulphate 0.7 g; caffeic acid 0.18 g; noble agar 20 g; ferric citrate solution 4.0 ml (2 mM)– was modified. The new medium, minimal synthetic caffeic acid medium (MSCAM), only contains caffeic acid and ferric citrate (caffeic acid 0.18 g/l; noble agar 20 g/l and 4 ml/l of ferric citrate solution).The medium was sterilized in an autoclave at 120 °C for 15 min. Twenty milliliters of the medium were dispensed in Petri plates under sterile conditions.

The following clinical and environmental isolates were studied: 170 human isolates of C. neoformans from the Adolfo Lutz Institute (Sao Paulo, Brazil) and from the Sezione Malattie Infettive (University of Torino, Torino, Italy), 10 C. neoformans isolates from bird droppings from the University of Messina (Messina, Italy), 10 Candida albicans strains from the Sezione Malattie Infettive (University of Torino), 10 Candida dubliniensis strains from the Medical Mycology Laboratory (Universidad del País Vasco, Bilbao, Spain), two Candida glabrata isolates, and one isolate each of Candida tropicalis and Saccharomyces cerevisiae from the Sezione Malattie Infettive (University of Torino). Identification of isolates was confirmed by conventional mycological methods such as colony and microscopic morphology, urease production, nitrate reduction, germ tube test, chlamydoconidium production on corn meal agar, and carbohydrate assimilation patterns using ATB ID 32 C kit (bioMérieux, France).

The isolates and strains were transferred onto fresh malt agar slants and incubated at 25 °C. After five days a loopful of the colonies was inoculated on MSCAM, and incubated at 37 °C in the dark for 4 h, and then for 24 h. Sterile normal CAAM was used as control. Each isolate was tested three times in triplicate. Depending on the brown pigment produced by each of the *C. neoformans* isolates tested after 4 h and 24 h of incubation at 37 °C, five groups were created: without pigment production (0), very poor pigment production (1), poor pigment production (2), good pigment production (3) and very good pigment production (4). The data obtained from the experiments performed three times in triplicate were evaluated by using the statistical Sign test.



Figure. Different pigment production by a C. neoformans isolate from an AIDS patient after 24 h incubation at 37 $^\circ\text{C}$ on the CAAM (left) and on the new MSCAM (right).

Results

All *C. neoformans* isolates produced brown pigment after 4 h and 24 h of incubation at 37 °C. After 4 h, 132 (73 %) *C. neoformans* isolates showed a poor pigmentation (categories 1 and 2) in the new MSCAM and 48 (27%) isolates showed highly-pigmented colonies (categories 3 and 4). At the same incubation time (4 h) using CAAM, 160 (89%) *C. neoformans* isolates showed a poor pigmentation of the colonies (categories 1 and 2) and 20 (11%) a notable pigmentation (categories 3 and 4) (Table). After 24 h of incubation, colonies from 146 (81%) strains showed a rich pigmentation (categories 3-4) in the new MSCAM medium and, on the contrary, only colonies from 85 (47%) *C. neoformans* strains showed a rich pigmentation (categories 3 and 4) in CAAM (Table).

In the new MSCAM after 4 h at 37 °C, 170 C. neoformans isolates showed higher pigment production (1.9 mean) than the same isolates tested in the normal CAAM (1.7 mean). The Sign test showed significant differences between both media (p = 0.006). It is interesting to outline that C. neoformans isolates were able to produce a low pigmentation in the new MSCAM after 2 h at 37 °C. On the contrary no pigmentation was observed in the normal CAAM (data not shown). After 24 h, C. neoformans isolates in the MSCAM produced higher pigment production (3.2 mean) than the same strains tested in the normal CAAM (2.5 mean) (Figure). The Sign test resulted very significant (p = 0.0001). No pigmentation was observed at 4 h and 24 h in any of the tested isolates from the species C. albicans, C. dubliniensis, C. glabrata, C. tropicalis and S. cerevisiae.

Table. Pi	gment production value	es of 180 Cryptococcus	neoformans strains tested	on MSCAM and CAAM after	4 h and 24 h incubation at 37 °C	С.
-----------	------------------------	------------------------	---------------------------	-------------------------	----------------------------------	----

		Incubation time					
		4 h		24 h			
Medium		1 and 2*	3 and 4*	1 and 2	3 and 4		
MSCAM	No. of isolates (%)	132 (73)	48 (27)	34 (19)	146 (81)		
CAAM	No. of isolates (%)	160 (68)	20 (11)	95 (53)	85 (47)		

*Pigmentation category: 1 and 2 = very poor and poor colony pigmentation; 3 and 4 = strong and very strong colony pigmentation.

Discussion

The only two compounds contained in the MSCAM allowed more rapid growth and pigmentation of colonies from *C. neoformans* isolates than the most common used CAAM. This new medium, MSCAM, is easy to prepare, store and costs less than CAAM. Absence of glucose in the new medium increases pigment production in *C. neoformans* clinical and environmental isolates. According to our results the absence of glucose does not interfere in *C. neoformans* phenoloxidase activity. It is also possible to hypothesize that bivalent or monovalent ions such as Ca⁺⁺, NH₄⁺⁺ or K⁺ do not interfere in *C. neoformans* phenoloxidase activity. According to Polacheck [16] reduction of ammonium sulphate does not interfere with melanin synthesis and consequently with *C. neoformans* pigmentation.

The concept of using differential media for isolating specific fungal pathogens is not new but problems encountered with differential media used for identification of *C. neoformans* have included a elevated cost, a complex medium preparation, and ill-defined interpretation. MSCAM is inexpensive and easy to prepare and store; moreover, it allows a rapid and clear identification of *C. neoformans* isolates from human and environmental sources.

> Authors thank Professor Marcia Melhem of the Adolfo Lutz Institute, Sao Paulo (Brazil) and professor Giuseppe Criseo of Messina University, Italy, for the C. neoformas strains. The project UPV 093.327-EB157/99 from the Universidad del País Vasco-Euskal Herriko Unibertsitatea granted part of this work.

References

- Chaskes S, Edberg SC, Singer JM. A DL-dopa drop test for the identification of *Cryptococcus neoformans*. Mycopathologia 1981; 74: 143-148.
- Chaskes S, Tyndall RL. Pigment production by *Cryptococcus neoformans* and other *Cryptococcus* species from aminophenols and diaminobenzenes. J Clin Microbiol 1978; 7: 146-152.
- Colom MF, Frasés S, Ferrer C, Martín-Mazuelos E, Hermoso de Mendoza M, Torres Rodríguez JM, Quindós G. Estudio epidemiológico de la criptococosis en España: primeros resultados. Rev Iberoam Micol 2001; 18: 99-104.
- Criseo G, Bolignano MS, De Leo F, Staib F. Evidence of canary droppings as an important reservoir of *Cryptococcus* neoformans. Zbl Bakt 1995; 282: 244-254.
- Denning DW, Stevens DA, Hamilton JR. Comparison of *Guizotia abyssinica* seed extract (Birdseed) agar with conventional media for selective identification of *Cryptococcus neoformans* in patients with Acquired immunodeficiency syndrome. J Clin Microbiol 1990; 28: 2565-2567.
- 6. Douchet C, Chandenier J, Barrabes A, Therizol-Ferly M, Lenoble R. Reconaissance rapide de colonies de *Cryptococcus neoformans*. J Mycol Med 1995; 5: 122-123.
- Edberg SC, Chaskes SJ, Alture-Werber E, Singer JM. Esculin-based medium for isolation and identification of *Cryptococcus neoformans*. J Clin Microbiol 1980; 12: 332-335.

- Fleming WH IIII, Hopkings JM, Land GA. New culture for the presumptive identification of *Candida albicans* and *Cryptococcus neoformans*. J Clin Microbiol 1977; 5: 236-243.
- Garau M, Del Palacio A. Artritis por Cryptococcus neoformans en receptor de trasplante renal. Rev Iberoam Micol 2002; 19: 186-189.
- Healy ME, Dillavou CL, Taylor G. Diagnostic medium containing inositol, urea and caffeic acid for selective growth of *Cryptococcus neoformans*. J Clin Microbiol 1977; 6: 387-391.
- Hopfer RL, Blank F. Caffeic acid containing medium for identification of *Cryptococcus neoformans*. J Clin Microbiol 1975; 2: 115-120.
- Kaufmann CS, Merz WG. Two rapid pigmentation tests for identification of *Cryptococcus neoformans*. J Clin Microbiol 1982; 15: 339-341.
- Kwon-Chung KJ, Rhodes JC. Encapsulation and melanin formation as indicators of virulence in *Cryptococcus neoformans*. Infect Immun 1986; 51: 218-223.
- Paliwal DK, Randhawa HS. A rapid pigmentation test for identification of *Cryptococcus neoformans*. Antonie van Leeuwenhoek 1978; 44: 243-246.

- Perfect JR, Casadevall A. Cryptococcosis. Infect Dis Clin North Am 2002; 16: 837-874.
- Polacheck I, Hearing VJ, Kwon-Chung KJ. Biochemical studies of phenoloxidase and utilization of cathecolamines in *Cryptococcus neoformans*. J Bacteriol 1982; 150: 1212-1220.
- Polak A. Melanin as a virulence factor in pathogenic fungi. Mycoses 1989; 33: 215-224.
- Rubio M, De Vroey C, Chalon E, Swinne D. An improved medium for the isolation of *Cryptococcus neoformans* from pigeon droppings. Sabouraudia 1984; 22: 345-346.
- Schønheyder H, Stenderup A. Isolation of Cryptococcus neoformans from pigeon manure on two media inducing pigment formation. Sabouraudia 1982; 20: 193-197.
- Vidotto V, Aoki S, Campanini G. A vitamin-free minimal synthetic medium for *Cryptococcus neoformans*. Mycopathologia 1996; 133: 139-142.