



Original article

## Synergism of voriconazole or itraconazole with other antifungal agents against species of *Fusarium*

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### ABSTRACT

**Background:** Infections caused by *Fusarium* are difficult to treat because these fungi show in vitro and in vivo resistance to practically all the antifungal agents available, which explains the high mortality rates. An attempt to overcome fungal resistance is the combination of antifungal agents, especially those with different mechanisms of action.

**Aims:** Evaluate the in vitro interactions of combinations of voriconazole or itraconazole with other antifungal agents against 32 isolates of *Fusarium* spp.: *Fusarium chlamydosporum*, *Fusarium oxysporum*, *Fusarium proliferatum* and *Fusarium solani*.

**Methods:** Drug interactions were assessed by a checkerboard microdilution method that also included the determination of the MIC of each drug alone according to CLSI (Clinical and Laboratory Standards Institute) document M38-A2, 2008.

**Results:** The best combinations were voriconazole+terbinafine which showed synergism against 84% of *Fusarium* strains. Other synergistic combinations were voriconazole+itraconazole (50%), voriconazole+fluconazole (50%), voriconazole+miconazole (38%), voriconazole+flucytosine (22%) and voriconazole+ketoconazole (25%). The synergisms observed with itraconazole combinations were: itraconazole+terbinafine (25%) and itraconazole+flucytosine (9.37%). The antagonisms observed were: voriconazole+fluconazole (3%) and itraconazole+flucytosine (12.5%).

**Conclusions:** The synergism showed by voriconazole+terbinafine was remarkable. To better elucidate the potential usefulness of our findings, new in vivo and in vitro studies deserve be performed.

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## Sinergia del voriconazol y el itraconazol con otros agentes antimicóticos frente a *Fusarium* spp.

### RESUMEN

**Antecedentes:** Las micosis causadas por *Fusarium* son difíciles de tratar porque tanto *in vivo* como *in vitro* estos hongos muestran resistencia a casi todos los fármacos antimicóticos disponibles, lo que explica las altas tasas de mortalidad. Una tentativa de resolver la resistencia fúngica es combinar los fármacos antifúngicos, en especial los preparados con mecanismos de acción diferentes.

**Objetivos:** Valorar las interacciones *in vitro* de la combinación de voriconazol o itraconazol con otros fármacos antimicóticos frente a 32 aislamientos de *Fusarium* spp: *Fusarium chlamydosporum*, *Fusarium oxysporum*, *Fusarium proliferatum* y *Fusarium solani*.

**Métodos:** Las interacciones farmacológicas se valoraron con el método de microdilución en tablero de ajedrez, que también incluyó la determinación de la concentración inhibitoria mínima de cada fármaco por separado de acuerdo con el documento M38-A2, 2008, del Clinical and Laboratory Standards Institute.

**Resultados:** Las mejores combinaciones fueron voriconazol+terbinafina que mostró sinergia frente al 84% de los aislamientos de *Fusarium*. Otras combinaciones sinérgicas fueron: voriconazol+itraconazol (50%), voriconazol+fluconazol (50%), voriconazol+miconazol (38%), voriconazol+flucitosina (22%) y voriconazol+ketoconazol (25%). Las sinergias observadas con las combinaciones de itraconazol fueron itraconazol+terbinafina (25%), e itraconazol+flucitosina (9,37%). Los antagonismos observados fueron voriconazol+fluconazol (3%) e itraconazol+flucitosina (12,5%).

#### Palabras clave:

*Fusarium* spp.

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**Conclusiones:** La sinergia observada con voriconazol + terbinafina fue extraordinaria. Para dilucidar mejor la utilidad potencial de los hallazgos del presente estudio, han de realizarse nuevos estudios tanto in vivo como in vitro.

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*Fusarium* species are a common soil saprophyte and important plant pathogens,<sup>13</sup> and they may occasionally cause infection in animals.<sup>15</sup> In humans, *Fusarium* species cause a broad spectrum of infections, including superficial, locally invasive, and disseminated disease.<sup>22</sup> The infections in severely immunocompromised patients are associated with a poor clinical response to antifungal therapy and high mortality rates.<sup>9,23,28</sup> So, the successful outcome is determined by the degree of immunosuppression and the extent of the infection.<sup>23,26</sup> Due to its lesser frequency than other mycosis, as aspergilosis and candidiasis, the optimal treatment of fusariosis has not been established.<sup>7,28</sup>

This scenario is usually worsened because *Fusarium* infections are relatively resistant to treatments with antifungal agents, also confirmed by the susceptibility tests.<sup>4,5,17,26</sup> In general *Fusarium* species are resistant to echinocandins whilst susceptibility to amphotericin B, voriconazole and posaconazole are variable.<sup>9</sup> Therefore the use of a drug alone to treat fusariosis has not changed the mortality rates, which has remained high among immunocompromised patients.<sup>7</sup> Combination therapy could be used for improving the efficacy of antimicrobial therapy for difficult-to-treat infections, expanding the antifungal activity.<sup>1,18</sup> Salvage therapy combining amphotericin B + voriconazole has been used with good response.<sup>9</sup>

In general, the in vitro studies with associations of antifungal agents combines agents from different classes but previous studies have also reported synergism between two azoles as fluconazole + miconazole<sup>20</sup> or fluconazole + itraconazole.<sup>31</sup> Here we also explored this idea but focusing on voriconazole combined with azoles, terbinafina and flucytosine against 32 *Fusarium* isolates. The effects of the combinations between itraconazole plus terbinafina or flucytosine were also investigated.

## Materials and methods

Thirty-two clinical *Fusarium* strains were tested in this study: *Fusarium chlamydosporum* (4), *Fusarium oxysporum* (8), *Fusarium proliferatum* (2), *Fusarium solani* (17) and *F. solani* ATCC 36031 (1). The strains were obtained from different sources, including blood culture ( $n=15$ ), tissue biopsy ( $n=10$ ), cornea ( $n=3$ ), sediment of dialysate from CAPD ( $n=2$ ), fissures in interdigital area ( $n=1$ ), and contact lens solution ( $n=1$ ). Isolation and identification of the isolates were performed by standard microbiological and molecular techniques. Molecular analysis was performed to confirm the identity of each *Fusarium* isolate. A DNA fragment comprising an internal transcribed spacer (ITS), which was amplified using primers ITS1 (5'-TCCGTAGGTGA-ACCTGCGG-3') and ITS4 (5'-TCCTCCGCTATTGAT-ATGC-3').<sup>25</sup> The amplified fragments were sequenced, and the sequences were compared with DNA sequences of *Fusarium* obtained from GenBank which accession numbers were the ones that follow: *F. chlamydosporum* (HQ696899, HQ696900, HQ696908, HQ696909), *F. oxysporum* (HQ696898, HQ696897, HQ696896, HQ696895, HQ696894, HQ696893, HQ696892, HQ696891), *F. proliferatum* (HQ696886, HQ696887), *F. solani* (HQ696874, HQ696875, HQ696876, HQ696877, HQ696878, HQ696879, HQ696880, HQ696881, HQ696882, HQ696883, HQ696884, HQ696885, HQ696901, HQ696902, HQ696903, HQ696904, HQ696905).

Antifungal drugs were obtained as standard powders of known potency. Voriconazole (VCZ) (Pfizer, Brazil), terbinafina (TBF) (Sigma Pharma, Brazil), itraconazole (ITZ) (Cristalia, Brazil), miconazole (MCZ) (Cristalia, Brazil), and ketoconazole (KTZ) (Cristalia, Brazil) were dissolved in dimethylsulfoxide (Sigma Chemical Co.) while fluconazole (FCZ) (Medley, Brazil) and flucytosine (FCY) (Hoffman La Roche, Switzerland) were dissolved in distilled sterile water to make stock solutions. Final dilutions were made in RPMI 1640 with glutamine and without sodium bicarbonate (Gibco BRL–Life Technologies) buffered to pH 7.0 with MOPS 0.165 M (Sigma Chemical Co.) and adjusted with 0.1 M NaOH. The final concentration ranges were 0.25–32 µg/mL for flucytosine, 0.5–64 µg/mL for fluconazole and 0.125–16 µg/mL for miconazole, ketoconazole, itraconazole, voriconazole and terbinafina.

Drug interactions were assessed by a checkerboard microdilution method that also included the determination of the minimum inhibitory concentration (MIC) of each drug alone according to CLSI M38-A2 (Clinical and Laboratory Standards Institute; CLSI, 2008)<sup>10</sup> recommendations with serial twofold dilutions of drug combination. *Candida parapsilosis* ATCC 22019, *Candida krusei* ATCC 6258 and *Aspergillus flavus* ATCC 204304 were included as quality control strains for the determination of the MICs.

The isolates were cultured on potato dextrose agar slants at 35 °C for 3 days and at 25 °C for the following 4 days. The inoculum was prepared by scraping the surface of 7 days old fungal colonies from the agar growth using a loop and suspending the material in sterile saline solution. The density of conidial suspension was adjusted to a 70% transmittance at 530 nm. After this, the suspensions were diluted 1:50, and then 1:2 when 100 µL were inoculated in the wells containing an equal volume of drugs.

Aliquots of 50 µL of drug A plus 50 µL of drug B at a concentration four times the targeted final concentration were dispensed in the wells in order to obtain a two-dimensional checkerboard. Conidial suspensions were prepared as described above, and 100 µL of the inoculum were added to the wells. The microtiter plates were incubated for 48 h at 35 °C. For each strain tested, a positive (inoculum diluted) and negative control (only RPMI) were performed.

The MIC endpoints for the antifungal agents alone were: (a) the lowest concentration of drug that produced complete inhibition (100%) of the fungal growth for voriconazole and itraconazole; (b) the lowest concentration that produced 80% of growth inhibition for tests with terbinafina and (c) the lowest concentration that inhibited 50% of the fungi growth for tests with flucytosine, ketoconazole and miconazole. As all combined MICs contained voriconazole or itraconazole, in the combinations tests the MICs were considered as the lowest concentration that produced 100% of growth inhibition. Off scale MICs were converted to the next-higher twofold concentration.

Drug interactions were evaluated using a checkerboard titration method for the following combinations: VCZ + TBF, VCZ + ITZ, VCZ + FCZ, VCZ + MCZ, VCZ + FCY, VCZ + KTZ, ITZ + TBF and ITZ + FCY. The fractional inhibitory concentration index (FICI) was used to classify drug interaction. The FICI was calculated using the following formula: (MIC of drug A in combination/MIC of drug A alone) + (MIC of drug B in combination/MIC of drug B alone). Drug interactions were considered as synergistic for FICI ≤ 0.5, not interactive (indifferent) when the FICI was between 0.5 and 4 and antagonistic for FICI > 4.<sup>18</sup>

**Table 1**  
Geometric mean fractional inhibitory concentration indexes (FICIs) for voriconazole and itraconazole in association with antifungal agents against *Fusarium* species.

<i>Fusarium</i> species	Voriconazole						Itraconazole	
	TBF	ITZ	FCZ	MCZ	KTZ	FCY	TBF	FCY
<i>F. chlamydosporum</i>	0.44	0.78	1.25	2.02	2.03	3.00	0.42	0.80
<i>F. nygamai</i>	0.19	0.56	1.00	1.01	2.00	2.00	0.40	0.48
<i>F. oxysporum</i>	0.27	0.69	0.88	0.88	0.88	1.50	0.62	3.4
<i>F. proliferatum</i>	0.17	0.33	0.59	1.02	1.05	1.25	0.44	0.56
<i>F. solani</i>	0.45	0.98	1.07	1.06	0.71	1.31	0.89	4.25
General FICI mean	0.39	0.86	1.07	1.05	1.01	1.41	0.81	3.9

FCZ, fluconazole; FCY, flucytosine; ITZ, itraconazole; KTZ, ketoconazole; MCZ, miconazole; TBF, terbinafine.

Drug interactions were classified as synergistic when the FICI was less than or equal to 0.5, indifferent when the FICI was greater than 0.5 and less than or equal to 4 and antagonistic when the FICI was greater than 4.

## Results

Total absence of activity against all *Fusarium* strains studied was observed for flucytosine (MIC > 32 µg/mL), fluconazole (MIC > 64 µg/mL) and itraconazole (MIC > 16 µg/mL) when tested alone. The susceptibility to the rest of the antifungal agents showed the following ranges: ketoconazole (2–6 µg/mL), miconazole (4–16 µg/mL), terbinafine (2–16 µg/mL) and voriconazole (2–8 µg/mL).

The results of the checkerboard analysis (FICI means) for voriconazole and itraconazole combinations against *Fusarium* strains are summarized in Table 1. The combination VCZ+TBF showed synergism for 27 strains (84%). Indifference was observed in one strain of *F. chlamydosporum*, one strain of *F. oxysporum* and three strains of *F. solani*, totaling 16% of indifferent interactions. Antagonisms were not observed for this combination. The interactions obtained by the combinations are shown in Table 2.

When VCZ was combined with ITZ, significant reductions in MIC geometric mean to ITZ (from >16 to 0.125 µg/mL) and in MIC geometric mean to VCZ (3.2–1.95 µg/mL) were present. This combination was synergistic for 50% of the strains and equally indifferent for all of them.

The mean MIC to FCZ for all isolates have decreased from >64 µg/mL to 0.125 µg/mL when it was used in combination with VCZ. Here, 53% of the strains tested demonstrated synergism, 44% were indifferent and against one isolate of *F. solani* this combination was antagonistic.

When VCZ was combined with MCZ the number of synergistic interactions was 12/32 (37.5%) and 62.5% were indifferent interactions but antagonism was not detected.

The combination VCZ plus FCY demonstrated synergism for only 7 isolates (22%) and the interactions were indifferent for 25 strains (78%). Although the MIC geometric mean for FCY dropped from >32 to 0.125 µg/mL when it was combined with the VCZ, this reduction was not significant to promote synergistic activity against the most *Fusarium* strains.

For the combination VCZ+KTC the predominance of indifference interactions (75%) were observed, while synergistic

**Table 2**  
Synergistic, indifferent and antagonistic interactions for combinations of voriconazole or Itraconazole with antifungal agents against 32 clinical *Fusarium* spp. strains.

Combinations of antifungal agents	Interactions n (%)		
	Synergism	Indifference	Antagonism
Voriconazole + terbinafine	27 (84%)	5 (16%)	0
Voriconazole + itraconazole	16 (50%)	16 (50%)	0
Voriconazole + fluconazole	17 (53%)	14 (44%)	1 (3%)
Voriconazole + miconazole	12 (37.5%)	20 (62.5%)	0
Voriconazole + flucytosine	7 (22%)	25 (78%)	0
Voriconazole + ketoconazole	8 (25%)	24 (75%)	0
Itraconazole + terbinafine	8 (25%)	24 (75%)	0
Itraconazole + flucytosine	3 (9.4%)	25 (78.1%)	4 (12.5%)

interactions were observed for 25% (8 of 32) of the strains tested. Synergy activity was observed for all *F. chlamydosporum*, *F. proliferatum*, and for only one *F. solani*; no one *F. oxysporum* strain was sensible for this combination as well as 94.1% of *F. solani* strains.

The best result for combinations with ITZ was ITZ+TBF which showed 8/32 (25%) of synergisms but no one antagonism. However, it is important to emphasize that this combination was indifferent for 100% of the *F. solani* strains and for 75% of the *F. oxysporum* strains.

With flucytosine the interactions showed 9.3% of synergism, 78.1% of indifference and 12.5% of antagonism. No one synergism was obtained from tests against *F. solani* and *F. oxysporum* which showed 11.7% and 25%, respectively of antagonistic interactions.

## Discussion

Combined antifungal therapy can be a potential strategy to improve the prognosis of some fungal infections. The in vitro studies evolving drugs associations have reported a lot of combinations including those with antifungal + non-antifungal agents, sometimes showing synergisms.<sup>1</sup> So, it seems rational to also explore meticulously the combinations antifungal + antifungal agents mainly against microorganisms difficult to treat as *Fusarium* species. In order to uncover potent antifungal combinations, the current study evaluated the interactions between VCZ with FCY, FCZ, ITZ, KTZ, MCZ and TBF and also the combinations evolving ITZ plus TBF or FCY against *Fusarium* strains by checkerboard microdilution assay.

In general our results showed a poor susceptibility of *Fusarium* for all the drugs tested alone, as previously described by some authors.<sup>4,5,26</sup> Despite the high in vitro MIC values, VCZ have been proposed as monotherapy for fusariosis in patients refractory to other drugs.<sup>9</sup> *F. proliferatum* demonstrated a major number of synergistic interactions while *F. solani*, and *F. oxysporum* showed a more resistant susceptibility pattern.

Combination between VCZ plus TBF showed the best synergism (84%) and antagonism was not observed for this combination. It is in accordance with Ortoneda et al.<sup>26</sup> and Córdoba et al.<sup>11</sup> who reported the same synergism in 72.7% and 72.4%, respectively. Azor et al.,<sup>5</sup> studying the susceptibility of 48 *Fusarium* strains against 11 antifungal agents, observed that terbinafine was the most active drug against all the species tested, what corroborates our results. The enhancement of the in vitro activity of VCZ when in combination with TBF has been explained by the blockage of ergosterol biosynthesis at different levels.<sup>11</sup>

In general the in vitro studies focusing associations of antifungal agents have combined drugs from different classes in order to add two mechanisms of action. So, the most studied combinations include polyene + azoles or polyene + pyrimidine analog (flucytosine) or azoles + echinocandins or polyene + echinocandins.<sup>18</sup> Combinations among antifungal agents plus non-antifungal agents have also been explored.<sup>1</sup> Our study has also included

combinations among azoles as VCZ + KTZ, VCZ + MCZ, and VCZ + ITZ. Azole–azole combinations have been employed on a limited number of studies.<sup>18</sup> The combination of FCZ plus ITZ in an experimental systemic cryptococcosis and meningitis resulted in improvements in tissue sterilization.<sup>31</sup> Another cornerstone for our investigation was Mykami et al.<sup>20</sup> who reported synergism between FCZ+MCZ against *Candida*. It was suggested that the mechanism of this effect would be the direct membrane damage induced by miconazole what might make the membrane more fragile and make easy the entrance of fluconazole into the fungal cell. Here we explored the same idea but focusing voriconazole in combination with other azoles. Among the azole + azole combinations tested, VCZ plus ITZ and VCZ plus FCZ have demonstrated 50% to 53% respectively of synergistic interactions against different species of *Fusarium* spp. The antagonistic interaction between VCZ plus FCZ (3%) have not been reported because this combination is not common; the mechanisms involved in these interactions seem to be unknown. Combinations of VCZ with MCZ and KTZ have demonstrated indifferent interactions for most strains tested. As far as we know combinations among voriconazole and azoles have not yet been studied against *Fusarium*.

The combinations azoles + FCY have been lesser studied than amphotericin B + FCY. Although previous investigations of the interactions between FCY with older azoles (miconazole and econazole) had suggested antagonism,<sup>14</sup> recent reports of triazole–FCY combinations have indicated synergism or indifference.<sup>2,6,16,21,24,30</sup> Here we obtained 22% of synergism by VCZ + FCY against *Fusarium*, in spite of indifferences being remarkable. Combination between VCZ and FCY has not yet been studied for *Fusarium*. The most combinations among azoles + FCY have been studied focusing *Cryptococcus* and *Candida* because these genera are primarily sensitive to FCY and the addition of a triazole suppresses the emergence of flucytosine-resistant mutants.<sup>18</sup>

The combination between ITZ and TBF showed 25% of synergism but no antagonism was registered. This is partially in accordance with the results reported by Ortoneda et al.<sup>26</sup> who found 36.3% of synergism and 45.4% of antagonism. This combination has been studied by other authors who have noticed synergisms against *Candida*,<sup>8</sup> *Aspergillus*,<sup>29</sup> and against two very problematic microorganism, as *Scedosporium prolificans*<sup>19</sup> and *Pythium insidiosum*.<sup>3</sup> The positive interaction may be due to the combined effect of TBF and the azoles on different targets in the ergosterol biosynthesis pathway.<sup>26</sup> Our results were similar to the finding pointed by Córdoba et al.<sup>11</sup> that this combination did not show synergism against *F. solani*, even though they have been detected for some isolates of *F. oxysporum*.

The combinations of FCY + ITZ or KTZ or posaconazole in animal models have resulted in improvements in survival<sup>12,27</sup> and tissue clearance<sup>31</sup> in cases of cryptococcal meningitis. In addition, in a clinical case series of systemic mycosis the outcomes with ITZ + FCY were comparable or better than those for ITZ alone.<sup>32</sup> The azole damages the fungal cell membrane, enabling increased uptake of flucytosine. This mechanism may explain potential synergism between azoles and flucytosine.<sup>18</sup> Here we showed only 3 (9.4%) synergistic interactions which were against *F. chlamydosporum* and *F. proliferatum* that confirm the variability of susceptibility among *Fusarium* species already suggested by others.<sup>11,23,26</sup>

Finally, our study demonstrated that the in vitro combination of VCZ against the genus *Fusarium* can be enhanced by other drugs showing different mechanisms of action including drugs of the same class as azoles. To better elucidate the potential usefulness of our findings, new in vivo and in vitro studies should be performed.

#### Conflict of interest

The authors declare that they have no conflict of interest.

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#### References

- Afeltra J, Vitale RG, Mouton JW, Verweij PE. Potent synergistic in vitro interaction between nonantimicrobial membrane-active compounds and itraconazole against clinical isolates of *Aspergillus fumigatus* resistant to itraconazole. *Antimicrob Agents Chemother*. 2004;48:1335–43.
- Allendoerfer RA, Marquis AJ, Rinaldi MG, Graybill JR. Combined therapy with fluconazole and flucytosine in murine cryptococcal meningitis. *Antimicrob Agents Chemother*. 1991;35:726–9.
- Argenta JS, Santurio JM, Alves SH, Pereira DIB, Cavalheiro AS, Spanemberg A, et al. In vitro activities of voriconazole, itraconazole and terbinafine alone or in combination against *Pythium insidiosum* isolates from Brazil. *Antimicrob Agents Chemother*. 2008;52:767–9.
- Arikan S, Lozano-Chiu M, Paetznick V, Nangia S, Rex JH. Microdilution susceptibility testing of amphotericin B, itraconazole, and voriconazole against clinical isolates of *Aspergillus* and *Fusarium* species. *J Clin Microbiol*. 1999;37:3946–51.
- Azor M, Gené J, Cano J, Manikandan P, Venkathay N, Guarro J. Less frequent *Fusarium* species of clinical interest: correlation between morphological and molecular identification and antifungal susceptibility. *J Clin Microbiol*. 2009;47:1463–8.
- Barchiesi F, Gallo D, Caselli F, Di Francesco LF, Arzeni D, Giacometti A, et al. In vitro interactions of itraconazole with flucytosine against clinical isolates of *Cryptococcus neoformans*. *J Antimicrob Chemother*. 1999;44:65–70.
- Campo M, Lewis RE, Kontoyiannis DP. Invasive fusariosis in patients with hematologic malignancies at a cancer center: 1998–2009. *J Infect*. 2010;60:331–7.
- Cantón E, Pemán J, Gobernado M, Viudes A, Espinel-Ingroff A. Synergistic activities of fluconazole and voriconazole with terbinafine against four *Candida* species determined by checkerboard, time-kill, and Etest methods. *Antimicrob Agents Chemother*. 2005;49:1593–6.
- Chen SCA, Playford EG, Sorrell T. Antifungal therapy in invasive fungal infections. *Curr Opin Pharmacol*. 2010;10:522–30.
- Clinical Laboratory Standards, Institute. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi. Approved standard. Document M38-A2. Wayne, PA: CLSI; 2008.
- Córdoba S, Rodero L, Vivot W, Abrantes R, Davel G, Vitale RG. In vitro interactions of antifungal agents against clinical isolates of *Fusarium* spp. *Int J Antimicrob Agents*. 2008;31:171–4.
- Craven PC, Graybill JR. Combination of oral flucytosine and ketoconazole as therapy for experimental cryptococcal meningitis. *J Infect Dis*. 1984;149:584–90.
- Dignani MC, Anaissie E. Human fusariosis. *Clin Microbiol Infect*. 2004;10:67–75.
- Dupont B, Drouhet E. In vitro synergy and antagonism of antifungal agents against yeast-like fungi. *Postgrad Med J*. 1979;55:683–6.
- Evans J, Levesque D, de Lahunta A, Jensen HE. Intracranial fusariosis: a novel cause of fungal meningoencephalitis in a dog. *Vet Pathol*. 2004;41:510–4.
- Ghannoum MA, Fu Y, Ibrahim AS, Mortara LA, Shafiq MC, Edwards JE, et al. In vitro determination of optimal antifungal combinations against *Cryptococcus neoformans* and *Candida albicans*. *Antimicrob Agents Chemother*. 1995;39:2459–65.
- Guarro J, Pujol I, Mayayo E. In vitro and in vivo experimental activities of antifungal agents against. *Antimicrob Agents Chemother*. 1999;43:1256–7.
- Johnson MD, MacDougall C, Ostrosky-Zeichner L, Perfect JR, Rex JH. Combination antifungal therapy. *Antimicrob Agents Chemother*. 2004;48:693–715.
- Meletiadi J, Mouton JW, Meis JFGM, Verweij PE. In vitro drug interaction modeling of combinations of azoles with terbinafine against clinical *Scedosporium prolificans* isolates. *Antimicrob Agents Chemother*. 2003;47:106–17.
- Mykami Y, Takahashi A, Yazawa K, Terao K, Ueno Y. Synergistic interaction of miconazole and fluconazole at sub-MIC level on *Candida albicans*. *Mycoses*. 1992;35:321–7.
- Nguyen MH, Barchiesi F, McGough DA, Lu VL, Rinaldi MG. In vitro evaluation of combination of fluconazole and flucytosine against *Cryptococcus neoformans* var. *neoformans*. *Antimicrob Agents Chemother*. 1995;39:1691–5.
- Nucci M, Anaissie E. Cutaneous infection by *Fusarium* species in healthy and immunocompromised hosts: implications for diagnosis and management. *Clin Infect Dis*. 2002;35:909–20.
- Nucci M, Anaissie E. *Fusarium* infections in immunocompromised patients. *Clin Microbiol Rev*. 2007;20:695–704.
- Odds FC. Interactions among amphotericin B, 5-fluorocytosine, ketoconazole, and miconazole against pathogenic fungi in vitro. *Antimicrob Agents Chemother*. 1982;22:763–70.
- O'donnell K, Sarver BA, Brandt M, Chang DC, Noble-Wang J, Park BJ, et al. Phylogenetic diversity and microsphere array-based genotyping of human pathogenic fusaria, including isolates from the multistate contact lens-associated US keratitis outbreaks of 2005 and 2006. *J Clin Microbiol*. 2007;45:2235–48.
- Ortoneda M, Capilla J, Javier Pastor F, Pujol I, Guarro J. In vitro interactions of licensed and novel antifungal drugs against *Fusarium* spp. *Diagn Microbiol Infect Dis*. 2004;48:69–71.

27. Polak A. Combination therapy of experimental candidiasis, cryptococcosis, aspergillosis and wangiellois in mice. *Chemotherapy*. 1987;33:381–95.
28. Raad II, Hachem RY, Herbrecht R, Graybill JR, Hare R, Corcoran G, et al. Posaconazole as salvage treatment for invasive fusariosis in patients with underlying hematologic malignancy and other conditions. *Clin Infect Dis*. 2006;42:1398–403.
29. Ryder NS, Leitner I. Synergistic interaction of terbinafine with triazoles or amphotericin B against *Aspergillus* species. *Med Mycol*. 2001;39:91–5.
30. Scheven M, Junemann K, Schramm H, Huhn W. Successful treatment of a *Candida albicans* sepsis with a combination of flucytosine and fluconazole. *Mycoses*. 1992;35:315–6.
31. Van Cutsem J. Therapy of experimental meningeal and disseminated cryptococcosis. *Mycoses*. 1993;36:357–67.
32. Viviani MA, Tortorano AM, Pagano A, Vigevani GM, Gubertini G, Cristina S, et al. European experience with itraconazole in systemic mycosis. *J Am Acad Dermatol*. 1990;23:587–93.